

### An Overview of a Novel Analytical Affinity Chromatography Column

### TSKgel<sup>®</sup> FcR-IIIA-NPR<sup>™</sup>

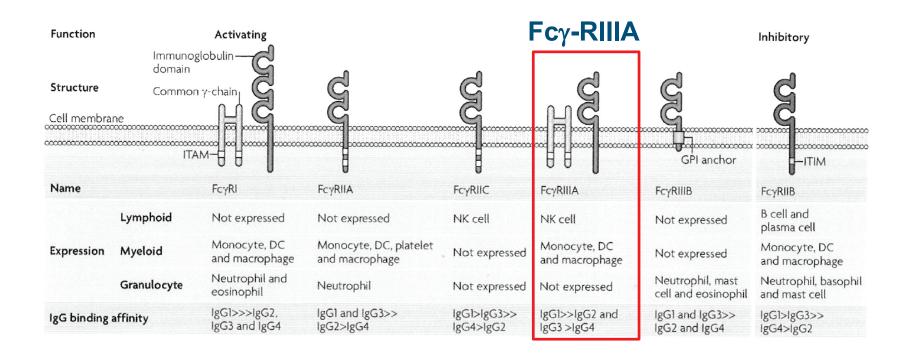
Atis Chakrabarti, Ph.D. Manager, Technical Service / Sr. Application Scientist Tosoh Bioscience LLC, King of Prussia, PA



- Introduction on development of novel FcR ligand
- Characteristics of the FcR column
- Applications
- Method Development aspects
- Conclusions

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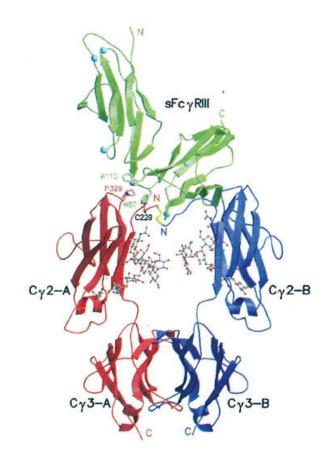




Kenneth G. C. Smith and Menna R. Clatworthy, Nature Reviews Immunology 10, 328-343 (May 2010)

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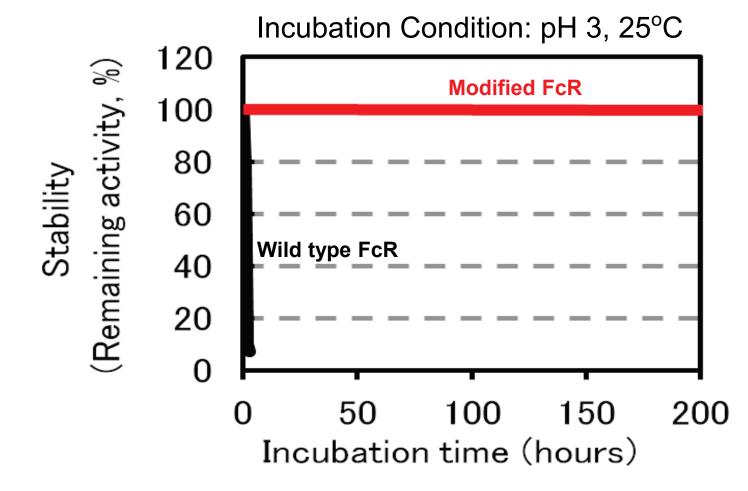
Modified Recombinant, 20 kDa

E. Coli expression system, non-glycosylated

Protein-based ligand

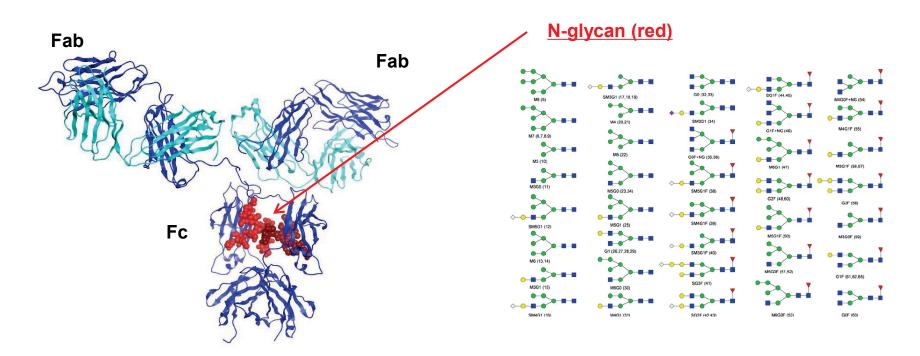
Ref: Peter Sondermann et al., Nature, vol. 406, 20 July 2000, 267-273





Stability of wild type FcyRIIIA is not enough for its use as affinity ligand



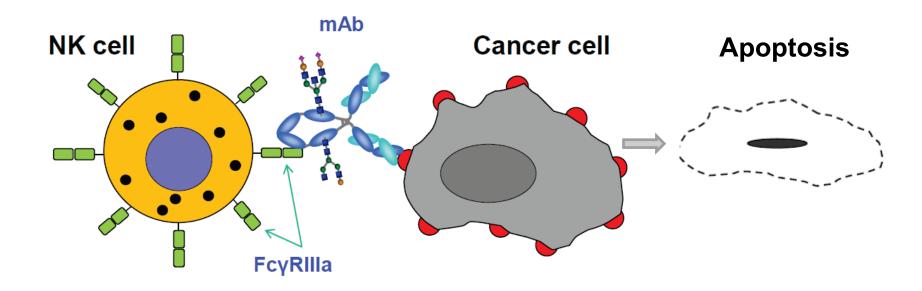


#### Structure of N-glycan attached to Asn297 of Fc region affects the drug efficacy

Ref:

- 1. Shinkawa T et al., J.Biol. Chem. 278, 3466-3473 (2003).
- 2. Pablo Umana et al., Nature Biotech. 17 FEB, 176-180 (1999).
- 3. Saphire, E.O et.al, Science 293:1155-1159 (2001)





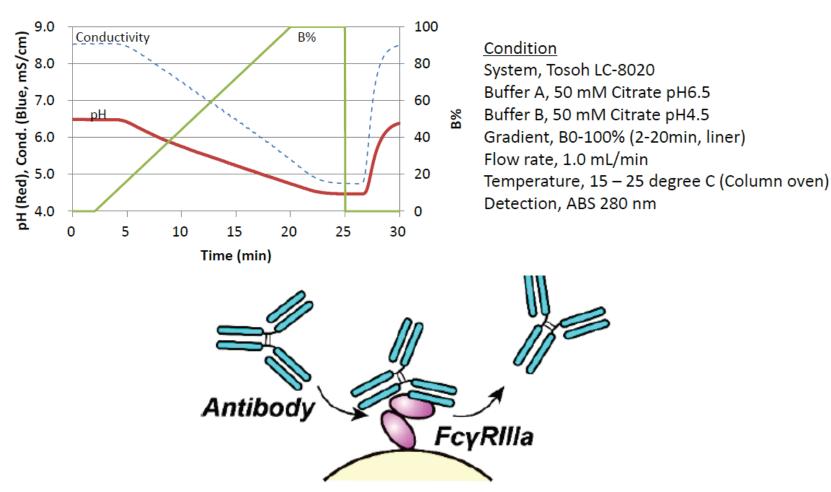
Types of N-Glycans at Asn297 of Fc receptor affect the binding affinity, thus ADCC activity

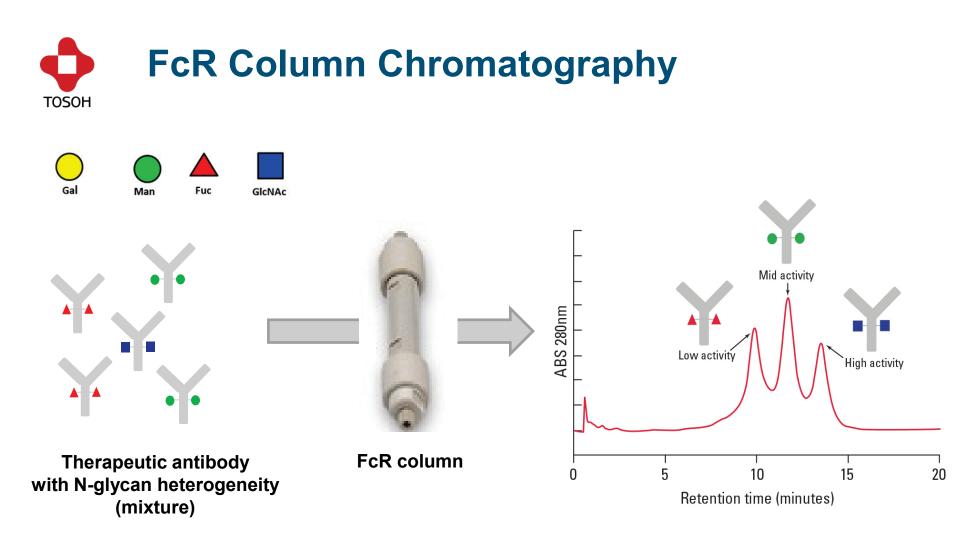
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Attributes		
Ligand	Modified recombinant FcγRIIIA ( <i>E. coli</i> expression system, non-glycosylated)	
Base matrix	nonporous resin, 5 μm	
Column	4.6 mm ID x 7.5 cm, PEEK	
Sample mass	5 – 50 μg of IgG (recommended)	
Standard flow rate	1.0 mL/min	
Temperature	15 °C - 25 °C	
pH range	4 - 8	
Recommended buffer system	A: 50 mmol/L citrate buffer, 150 mmol/L NaCl, pH 6.5 B: 50 mmol/L citrate buffer, 150 mmol/L NaCl, pH 4.5	
Maximum pressure	9 MPa	

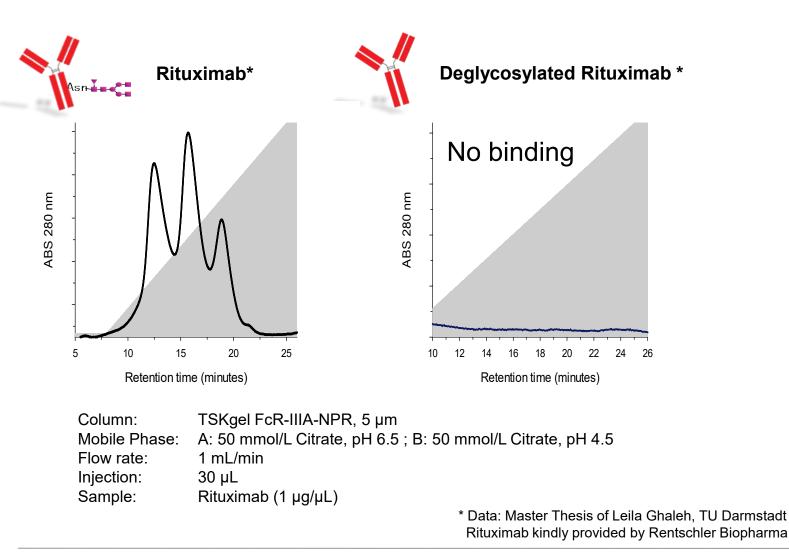
# **Standard Chromatographic Condition**





mAb components are separated based on the correlation between N-glycan structure and ADCC activity.

### Removal of N-Glycans Prevent mAb Binding to FcR Receptor



TOSOH



#### Modified FcyRIIIA Protein A 120 120 With 100 100 N-glycan 80 80 RU RU 60 60 Without Without 40 40 N-glycan N-glycan

0 0 200 400 600 0 0 200 400 600 Time (sec.) Time (sec.)

20

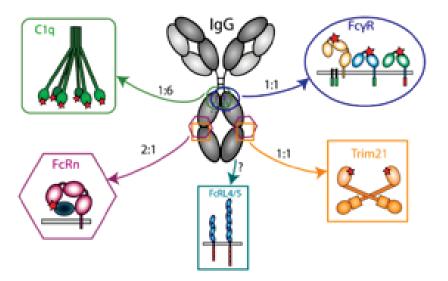
The modified FcR ligand does not bind to mAb without N-glycan.

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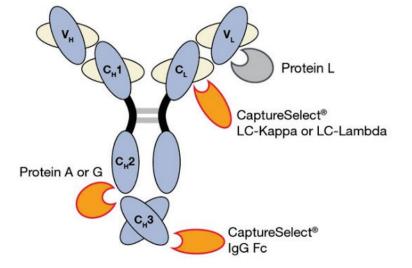
With

N-glycan





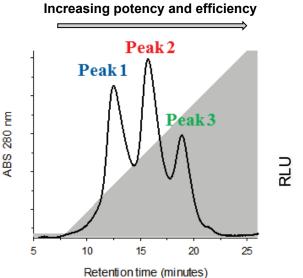
Ref: 1. Antibodies 2019, 8(2), 30; https://doi.org/10.3390/antib8020030



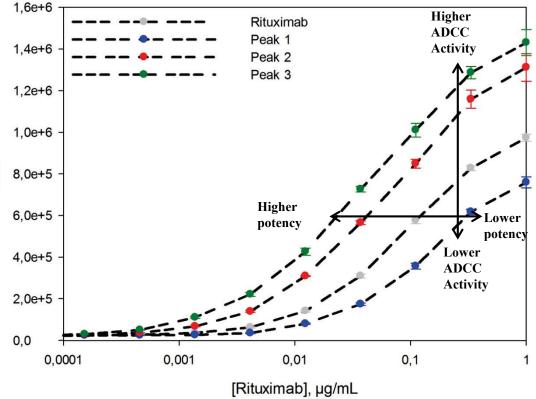
Ref: 2: Thermofisher.com

### $Fc\gamma$ receptor binding site is located near the hinge region of IgG





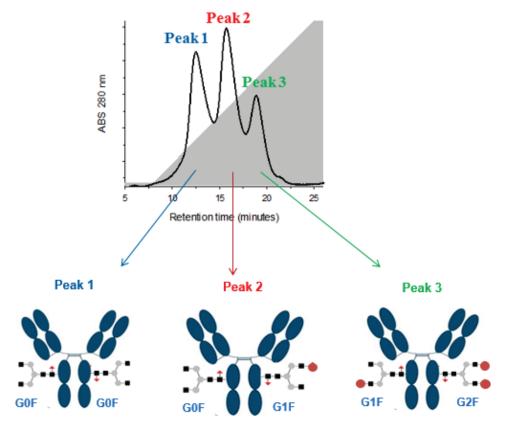
Antibody	EC <sub>50</sub> (µg/ mL)
Rituximab	0.098
Peak 1	0.153
Peak 2	0.072
Peak 3	0.049



### ADCC reporter bioassay (Promega)

\* Data: Master Thesis of Leila Ghaleh, TU Darmstadt Rituximab kindly provided by Rentschler Biopharma

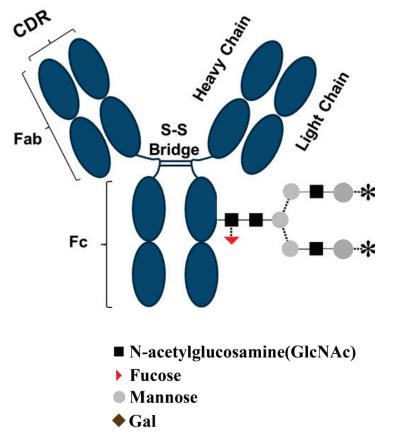


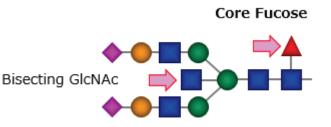


## The larger the number of terminal galactose, the higher the binding affinity.

\* Data: Master Thesis of Leila Ghaleh, TU Darmstadt Rituximab kindly provided by Rentschler Biopharma



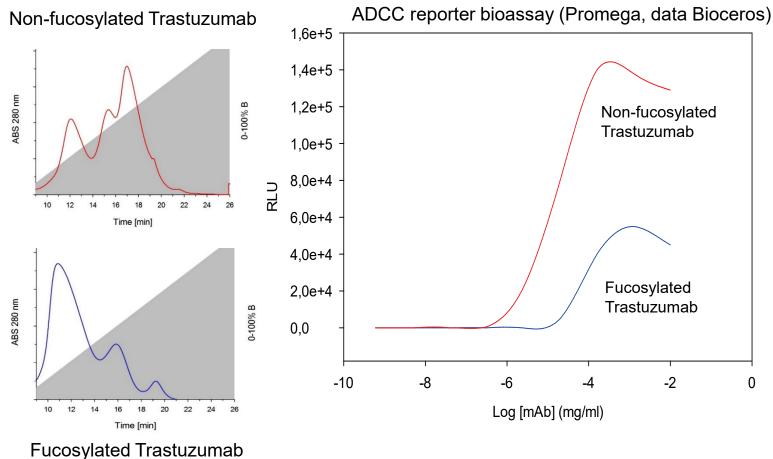




\*NeuNAc

#### Core fucose decreases ADCC activity

# Impact of Fucosylation



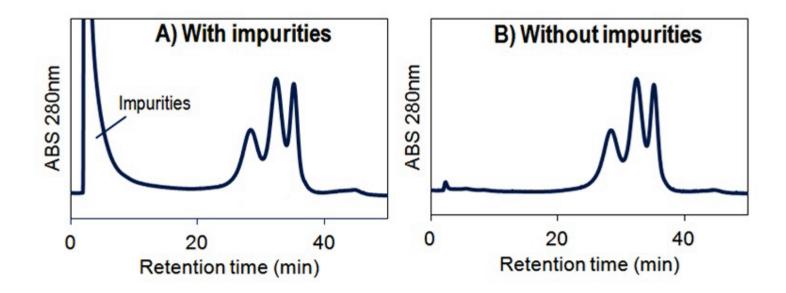
### ADCC activity correlates to the observed FcR affinity

\*Data: Master Thesis of Leila Ghaleh, TU Darmstadt \*\*Fucosylated and non-fucosylated Trastuzumab provided by Bioceros



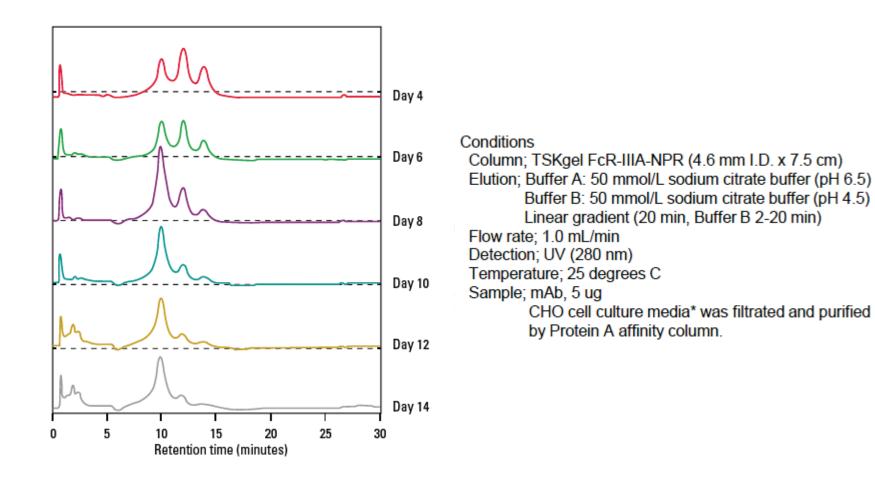
## **Applications of TSKgel FcR-IIIA-NPR**





- Host cell proteins don't affect the peak profile
- Only 5 µg of mAb without pre-processing is enough for analysis to monitor phases of development and production

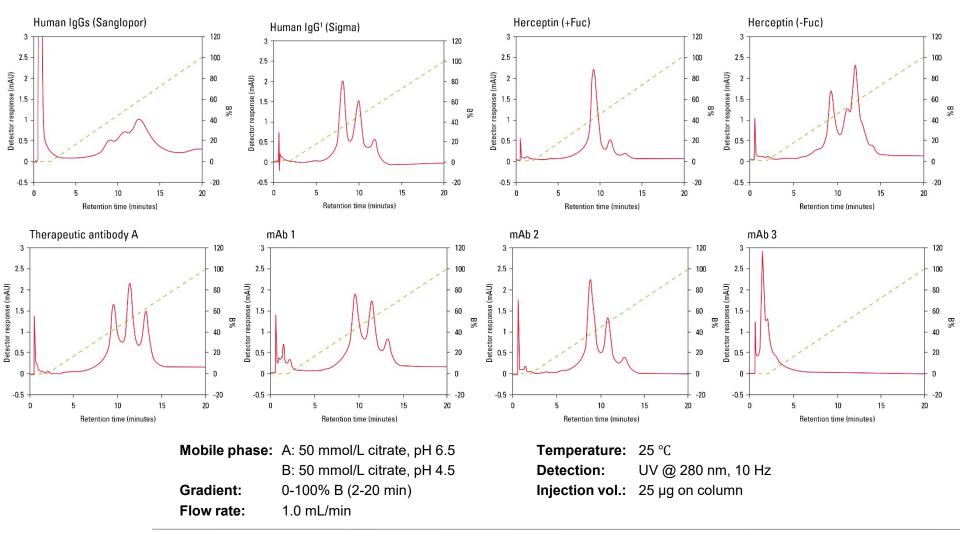
### Application 2: CHO Cell Culture Screening – Upstream Monitoring



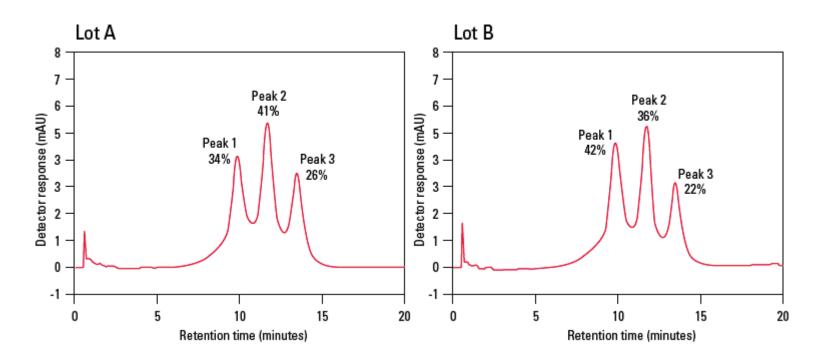
Antibody peaks shifted elution volume toward earlier elution, which suggests number of galactose in Fc glycan decreased with increased days of fermentation and suggests lower ADCC activity.

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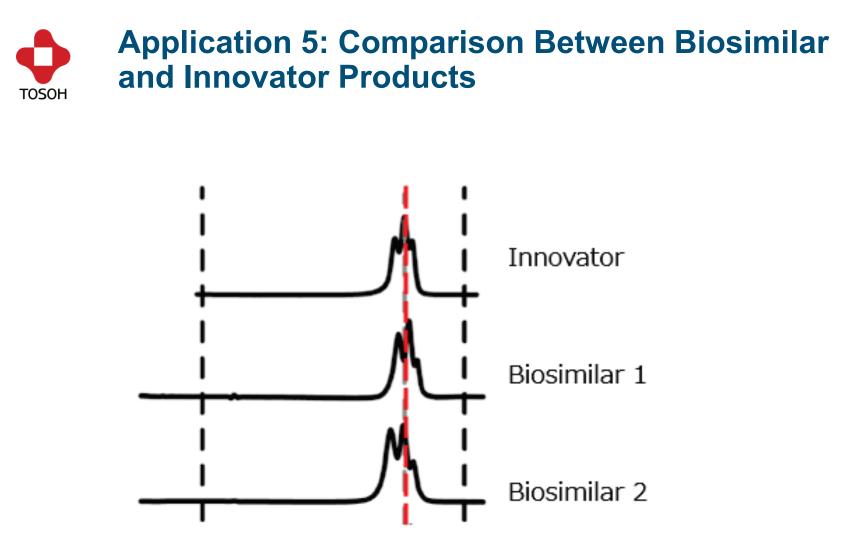






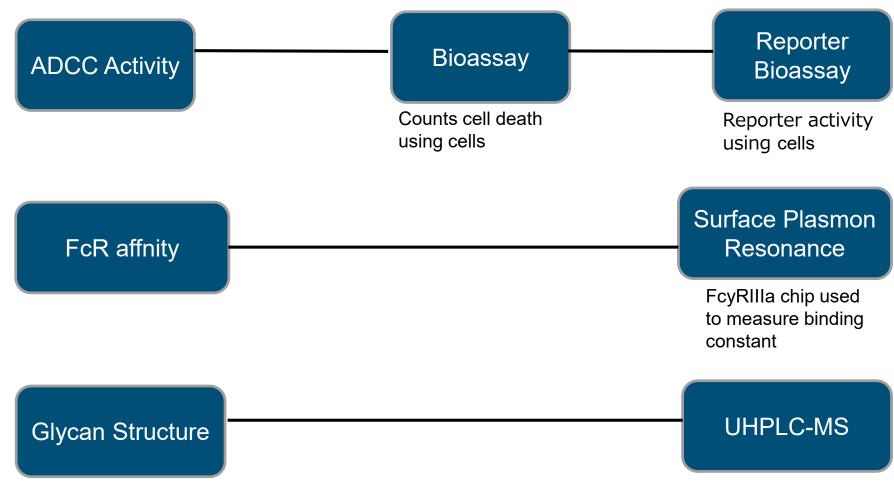


- N-glycan heterogeneity is main issue in quality control
- The TSKgel FcR-IIIA-NPR analytical column can be used for quality control of a therapeutic antibody.



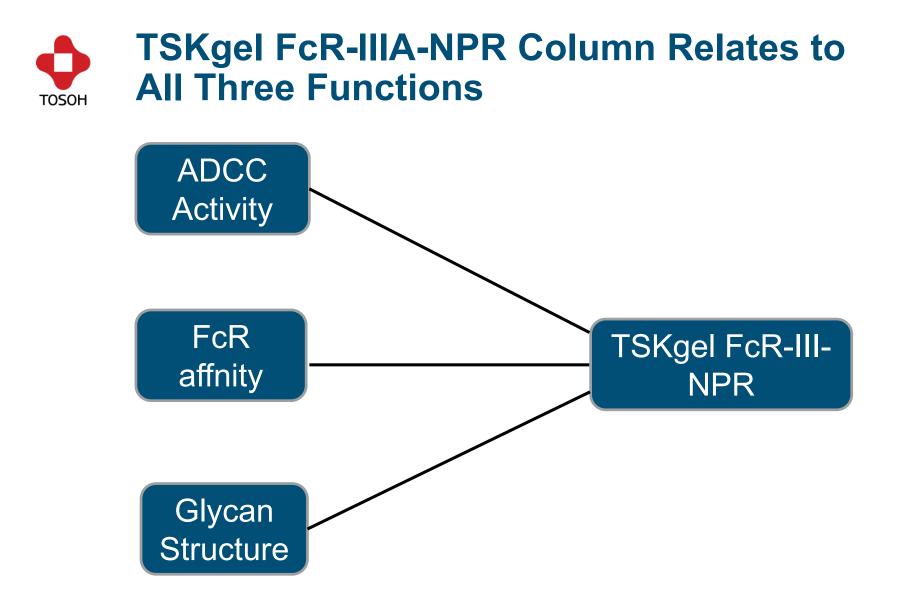
Ref.; M. Kiyoshi et al., Regulatory Science 2017, poster





#### Three different routes available for three different functions

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TSKgel FcR-IIIA-NPR column can be used for all three different functions



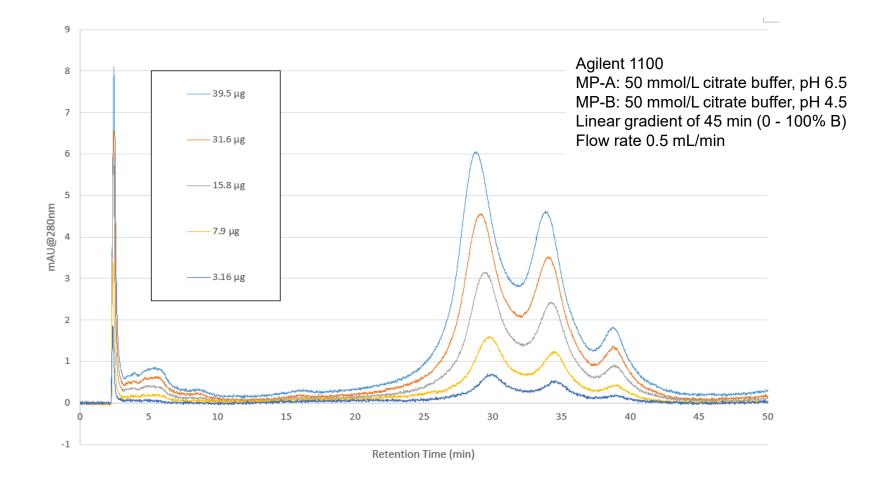
### Summary:

- 1. To monitor phases of development and production
- 2. Analysis of purified protein / mAb
- 3. Upstream monitoring of cell culture supernatant
- 4. Glycosylation profiling
- 5. QC analysis of lot-to-lot differences for mAb products
- 6. CHO cell line screening / to monitor fermentation stage of cell culture media
- 7. Comparison between biosimilar and innovator products
- 8. To correlate with ADCC activity

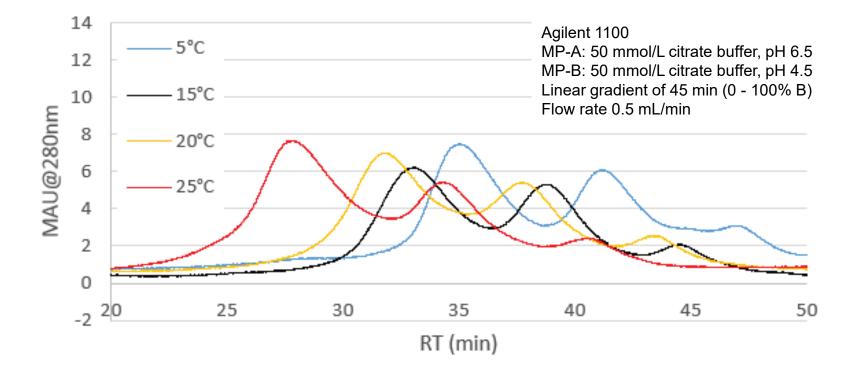


### **Method Development**

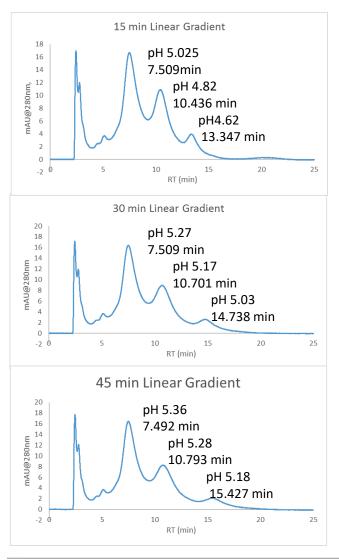
### Effect of Load (μg) of mAb on the Separation of the Three Glycoforms







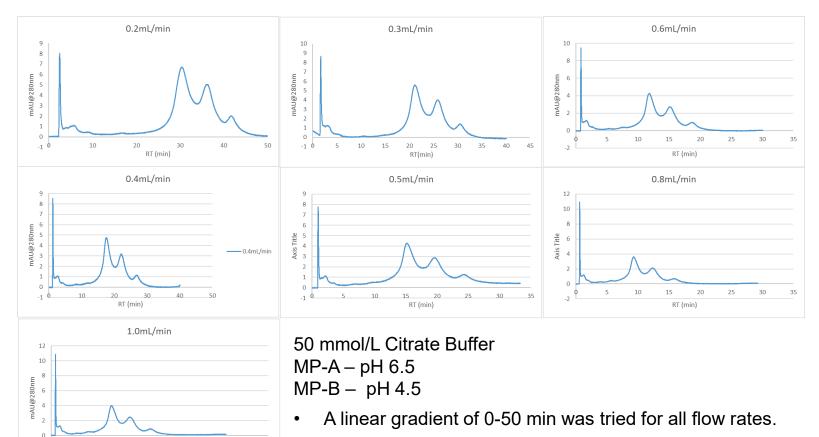
# **Effect of Gradient Slope on the Resolution**



Agilent 1200

MP-A: 50 mmol/L citrate buffer, pH 5.53 MP-B: 50 mmol/L citrate buffer, pH 4.5 Linear gradient over 15, 30 and 45 minutes Flow rate 0.2 mL/min





Peaks eluted roughly between pH 5.5 and 4.0.

### All the 3 glycoform peaks eluted by 67% B with approximately same % recovery

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RT (min)

10

5

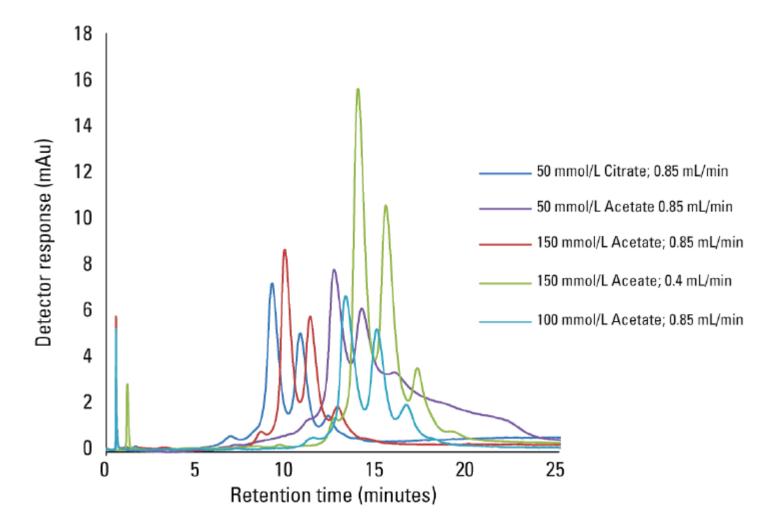
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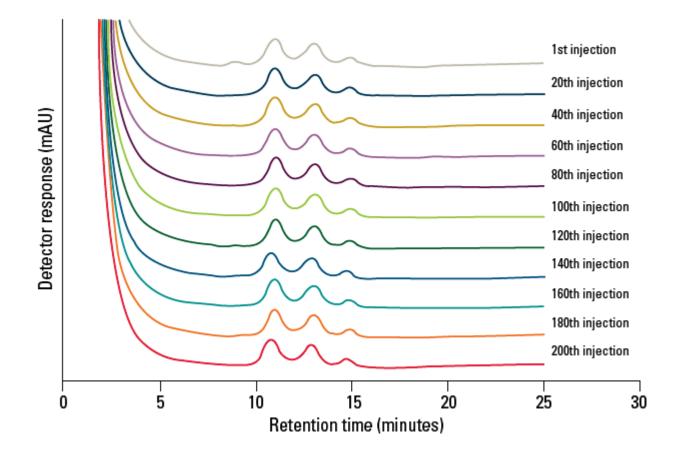
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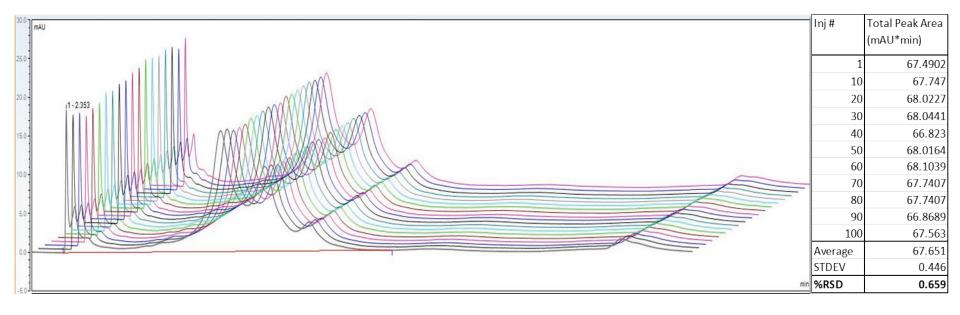




#### **CHO cell culture Supernatant**









## Conclusions



- The TSKgel FcR-IIIA-NPR column offers an easy and rapid analysis method for therapeutic antibodies.
- A qualitative analysis of the TSKgel FcR-IIIA-NPR stationary phase confirms the elution profile and order mimic the results of cell based ADCC assay.
- The stationary phase is selective for N-glycosylation of the Fc region, which may be useful in other assessments of PTMs and primary structure.
- The TSKgel FcR-IIIA-NPR column is useful for quality control and process analysis.