

# Characterization of Recombinant Protein Biotherapeutic using TSKgel® UP-SW2000 Size Exclusion Chromatography Column In-Line with LenS3® MALS Detector

## Introduction

Biotherapeutics are generally larger molecules such as peptides, proteins and monoclonal antibodies with monomer molecular weight ranging from 3,000-150,000 Da. The drug must remain free from impurities such as fragment, dimer and other higher order aggregates as they may cause severe immunogenic response. This becomes even more important particularly if the biotherapeutic protein is thermally susceptible. Historically, size exclusion is the preferred mode of chromatography used for separation and characterization of such applications. Here we report the online detection of absolute molecular weight of two recombinant protein samples using a 2  $\mu$ m size exclusion chromatography (SEC) column directly connected to the LenS3 Multi-Angle Light Scattering (MALS) detector.

## Materials and Methods

Samples: BSA (Calibration standard)  
Sample 1 – Recombinant Proteins (~90 kDa) at 1.72 mg/mL in mobile phase  
Sample 2 – Recombinant Proteins (~90 kDa) at 3.64 mg/mL in mobile phase

The samples were stored at -20 °C and thawed to 8 °C just before analysis. The concentration was adjusted by diluting the sample in mobile phase pre-chilled at 8 °C.

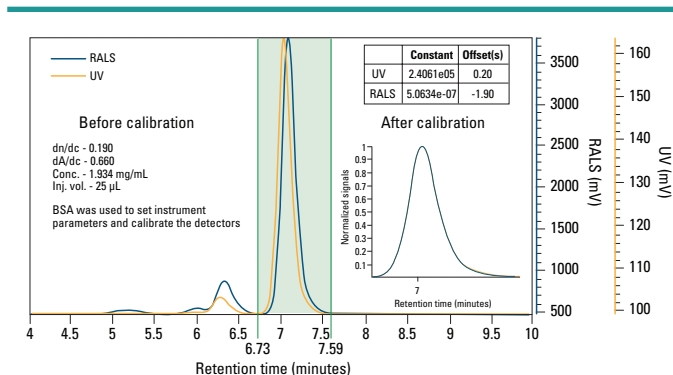
## Chromatographic Conditions

Instrument: ThermoFisher Ultimate® 3000 UHPLC and Chromeleon® software  
Column: TSKgel UP-SW2000, 2  $\mu$ m, 4.6 mm ID  $\times$  30 cm  
Mobile phase: BupH modified Dulbecco's phosphate buffer prepared in light scattering grade water and filtered through a 0.1  $\mu$ m PES membrane (The buffer was prepared from saline packs as per direction: Thermo Scientific – catalog # 28374, lot TL275790)  
Flow rate: 0.20 mL/min  
Detectors: UltiMate 3000 VWD variable wavelength detectors @ 280 nm wavelength and Tosoh LenS3 MALS detector (positioned in series: Column  $\rightarrow$  UV  $\rightarrow$  MALS)  
Column oven temp.: 25 °C  
Autosampler temp.: 8 °C  
Injection vol.: Sample 1: 25  $\mu$ L  
Sample 2: 10  $\mu$ L

## Results and Discussion

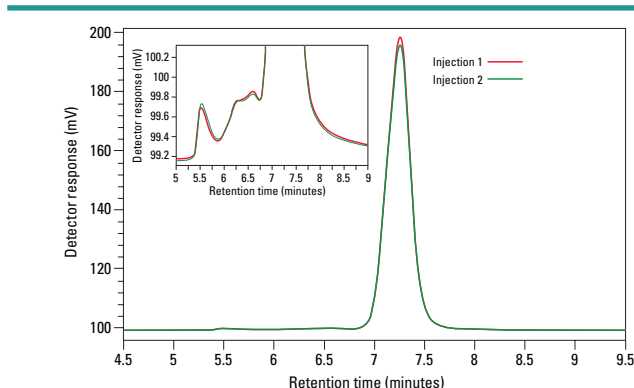
The multi-detector setup was calibrated using a freshly prepared bovine serum albumin (BSA) solution. **Figure 1** shows the overlay trace of UV and MALS detectors. The one-step calibration procedure in the SECview® software adjusted the dead volume between the detectors and corrected for the band-broadening effect caused by the in-series detector configuration while determining the detectors' calibration constants and offsets.

**Figure 1.** Calibration of the detectors using BSA protein standard

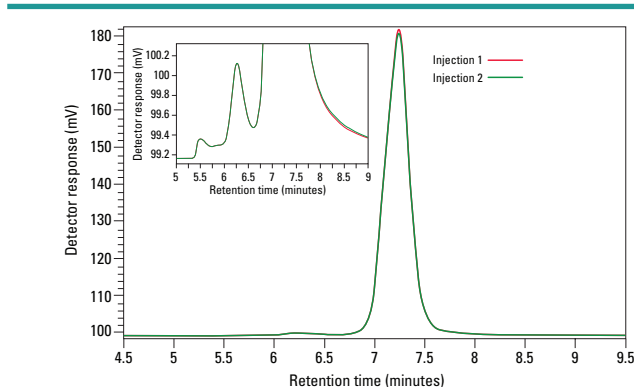


**Figures 2 and 3** illustrate the UV detector overlays for two consecutive injections of Sample 1 and Sample 2 respectively. Zoomed-in figures (in set) show the excellent separation resolution between the monomer and the aggregates obtained from TSKgel UP-SW2000 column.

**Figure 2.** UV detector overlay for Sample 1 - two consecutive injections

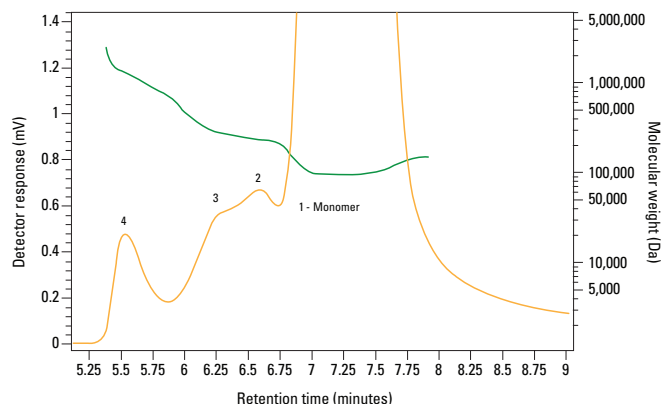


**Figure 3.** UV detector overlay for Sample 2 - two consecutive injections

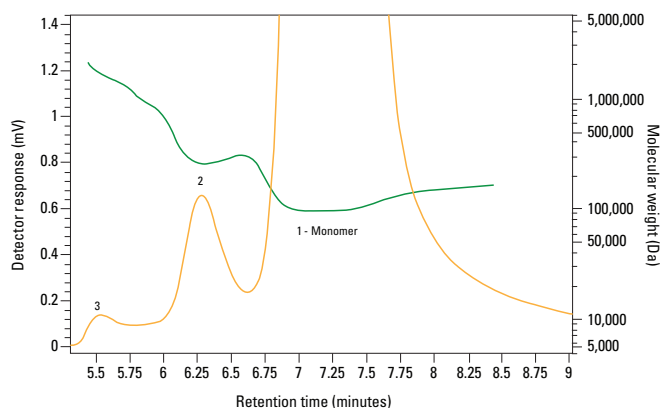


**Figures 4 and 5** demonstrate the molecular weight (green) profiles for Sample 1 and Sample 2 respectively. The concentrations of the aggregate contents differ in the two samples.

**Figure 4.** Molecular weight profile for Sample 1

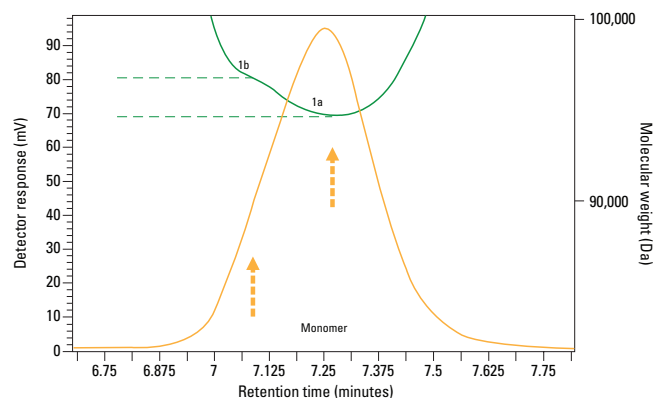


**Figure 5.** Molecular weight profile for Sample 2

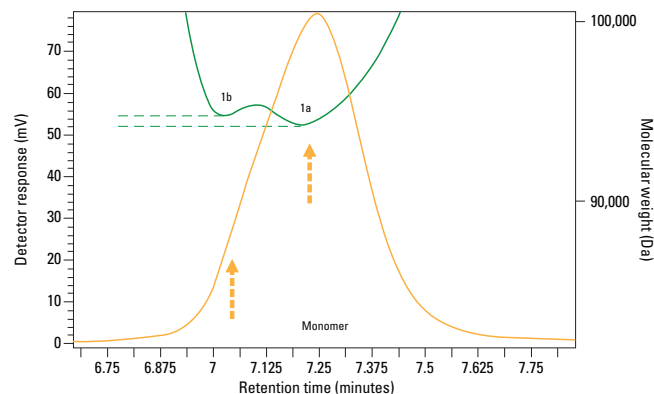


Looking closer at the UV trace, it appears that the monomer peaks in both samples illustrate a slight shoulder on the higher retention region, suggesting a bimodal shape. Further analysis using the molecular weight trace by the MALS detector reveals two separate populations of molecular weights. **Figures 6 and 7** zoom in on the monomer peaks and demonstrate the two molecular weight plateaus, 1a and 1b, in both samples. Considering the sensitive nature of these recombinant proteins to the ambient conditions, the shoulder peak (1b) suggests the beginning of temperature-induced modifications, which varies in extent in both samples.

**Figure 6.** Molecular weight profile of the monomer peak for Sample 1



**Figure 7.** Molecular weight profile of the monomer peak for Sample 2



**Tables 1 and 2** list the results of the chromatography analysis, including molecular weight and percent content, for the identified aggregate peaks.

**Table 1.** Results table for Sample 1

Peak	Retention Time*	Peak MW	% UV Area
1a	7.220	93,564 Da	98.06%
1b	-	95,811 Da	
2	6.593	229,024 Da	0.69%
3	6.355	276,217 Da	0.73%
4	5.533	1,285,996 Da	0.51%

**Table 2.** Results table for Sample 2

Peak	Retention Time*	Peak MW	% UV Area
1a	7.242	93,106 Da	98.73%
1b	-	93,602 Da	
2	6.282	251,078 Da	1.04%
3	5.540	1,631,227 Da	0.23%

\* Retention Times are reported in minutes

## Conclusions

This study shows that the molecular weight species including monomer and multiple aggregate levels present in the recombinant protein therapeutics could be determined and quantified using a SEC-MALS configuration. The TSKgel UP-SW2000 demonstrates excellent separation of the higher order aggregates from the monomer in both Sample 1 and Sample 2, as well as the slightly higher molecular weight temperature-induced variants that almost co-elute with the monomer. The column yields reproducible results, allowing accurate and precise chromatograms to be analyzed using the LenS<sub>3</sub> MALS detector. The LenS<sub>3</sub> accompanied by the SECview software also produces reproducible, accurate results in terms of MW determination for all peaks, in addition to area calculations, even at extremely low concentrations/presence of the aggregates.

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